



# PRIN Project Prot. 20173X8WA4

FIBRES: a multidisciplinary *minerealogical, crystal-chemical* and *biological* project to amend the paradigm of toxicity and cancerogenicity of mineral fibers.

### **Mineral Fibres:**



#### CROCIDOLITE

(UICC STANDARD)



#### **CHRYSOTILE**

(from Russia, « Yasnyi»)



#### **WOLLASTONITE NYAD G**

from Willsboro-Lewis (NY, USA)

### Citotoxicity Assessment on different in vitro models:

### **Cell monocultures**:

- I. Human monocyte and differentiated macrophage cells **THP-1**;
- II. Human Endothelial cells HECV
  - Cell Coculture HECV + THP-1 MO
- III. Human Foreskin Fetal Fibroblasts HFFF2

>3D Tissue Model **Epiairway™** (Mattek Corporation)

Permeability test on a synthetic pulmonary mucus L-Mu<sup>3</sup>Gel (Bac3Gel<sup>®</sup>Lda)

## Set-up of naive **THP-1** colture in indirect exposure to fibres



### MTS viability test at 48 and 72 hrs

#### THP-1 48\_72h



# Evaluation of early and late Apoptosis Cell Death after 48 hrs of fiber treatments (50 $\mu$ g/ml)



#### Magnification 20x; **Untreated**



#### Magnification 20x; **Chrysotile > 5 μm**



#### Magnification 20x; **Chrysotile < 5 μm**



#### Magnification 20x; Wollastonite

# Evaluation of early and late Apoptosis Cell Death after 72 hrs of fiber treatments (50 $\mu$ g/ml)



#### Magnification 20x; Untreated







#### Magnification 20x; Chrysotile > 5 μm





#### Magnification 20x; Wollastonite

## **Oxidative stress Analysis** ROS levels after 6 hrs exposure at 50 µg/ml





#### Directly exposed to the fibres



#### Assessment of HECV viability by DNA Assay

#### **HECV DNA assay**



\*p< 0.05 vs UT;

# Evaluation of early and late Apoptosis Cell Death after 48 hrs of fiber treatments (50 $\mu$ g/ml)



Magnification 60x; Untreated





Magnification 60x; Chrysotile > 5 μm









Magnification 60x; **Wollastonite** 

# Remarks.1

I. In THP-1 cells mechanisms of programmed death have been activated after 48 and 72 hrs. Although the ROS increase is not significant, a marked signal of DNA damage caused by both CHRs (< 5 and > 5  $\mu$ m) was observed.

II. The HECV cell viability decreased after 24 and 48 hrs in particular those treated with CHR> 5µm. In fact, as shown by LDH and Annexin/PI tests CHR > 5 µm leads to HECV cells death after 48 hrs. The H2AX protein phosphorylation was observed after 24 hrs of CHR > 5 µm treatment related to an increase of oxidative stress.



The human fibroblast line HFF2 was treated with HECV and THP-1 conditioned medium (CM) both subjected to previous treatments with mineral fibres, in order to evaluate the potential modulation and triggering of the inflammation in the cell layers involved in the carcinogenic process.

In this regard the mitochondrial activity of fibroblasts was evaluated after 72 hrs of treatments with CM by MTT test and staining with fluorescent dye JC-1.



Evaluation of HFF2 mitochondrial activity after 72 hrs exposure to HECV and THP-1 conditioned cell medium treatment



### JC-1 Protein Analysis by Confocal Microscopy

#### JC-1 72h\_t.c. HECV

UT











CTRL



CHR > 5 μm



\*\*p<0.01 vs UT; §§§§p<0.0001; §§§p<0.001 vs treated cells

# Remarks.3

Fiber-treated HECV CM affected the HFFF2 viability and the mitochondrial health. As indicated by confocal microscopy JC-1, producing red fluorescent «J-aggregates», at lower internal mitochondrial concentrations or low membrane potential is present as monomers, i.e. green J-monomers. This phenomenon is associated with programmed cell death (apoptosis).

# **3D Tissue Model EpiAirway™**



3D mucociliary tissue model consisting of normal, human-derived tracheal / bronchial epithelial cells (NHBE) from healthy / non-smoker donors.

Accumulated mucus by cells was removed from apical surface before the fiber treatments.



# Cytokine Analysis of IL-1α, IL-1β, TNF-α in EpiAirway culture medium after 24 and 48 hrs (MILLIPLEX<sup>®</sup> MAP system)



\*p<0.05,\*\*p<0.01,\*\*\*p<0.001,\*\*\*\*p<0.0001 vs UT

Cytokines were normalized to the percentage of viability measured by MTT test.

# Cytokine Analysis of IL-6,IL-8 in EpiAirway colture medium after 24 and 48 hrs

(MILLIPLEX<sup>®</sup> MAP system)



\*p<0.05,\*\*\*p<0.001,\*\*\*\*p<0.0001 vs UT

Cytokines were normalized to the percentage of viability measured by MTT test.

# Remarks.4

Both CHRs decreased the Epiairway tissue viability after 24 hrs more than CRO. At 48 hrs CRO-treated tissue reached the same level of CHRs. As well as CHRs promoted a greater release of TNF –  $\alpha$ , IL-1 $\beta$ , IL-6 and IL-8 cytokines especially after 48 hrs. A significative DNA damage was detected only in CHR > 5 – treated Epiairway.

## Work in progress Permeability test on synthetic pulmonary muci (Bac3Gel<sup>®</sup> Lda)

Mucus ~95% water

5% comprising electrolytes, lipids, DNA fragments, and proteins.

**Mucins** are HMW glycoproteins, consisting of a peptide backbone to which a huge amount of carbohydrate chains are attached. They permit the exchange of nutrients, water, gases, and hormones while being impermeable to most bacteria and pathogens<sup>1</sup>.

BAC 3GEL

**L-Mu<sup>3</sup>Gel** is a healthy lung mucus model with **MUCIN** capable of mimicking the steric and chemical barrier in physiologic conditions. (20 mg/ml porcine gastric mucin; 4.78 mg/ml sodium chloride)

**CF-Mu<sup>3</sup>Gel** is a pathologic lung mucus model with **MUCIN** capable of mimicking the steric and chemical barrier in pathologic conditions. (25.0 mg/ml porcine gastric mucin; 7.07 mg/ml sodium chloride)

## Mucus Mu<sup>3</sup>Gel Model set-up





Thickness of airway mucus in physiological conditions = 10 μm = 0.010 mm

given that the Transwell® support has a radius of 3.54 mm

Volume of L-Mu<sup>3</sup>Gel to introduce on the Transwell<sup>®</sup> support = 0.390 mm<sup>3</sup> = 0.390  $\mu$ l We are not able to pipet this volume!



Thickness of airway mucus in pathological conditions = 220 μm = 0.220 mm

given that the Transwell<sup>®</sup> support has a radius of 3.54 mm

Volume of Path,L-Mu<sup>3</sup>Gel to introduce on the Transwell<sup>®</sup> support = 8.66 mm<sup>3</sup> = 8.66  $\mu$ l  $\cong$  10  $\mu$ l

## In vitro set – up Mu<sup>3</sup>Gel Model





The medium placed on the basolateral side of the Thincert<sup>®</sup> insert was collected and stored at -20°C to analyze the content of metal ions crossing the mucus layer by ICP-MS.

### L-Mu3Gel – Healthy lung mucus

48h

72h



### CF-Mu<sup>3</sup>Gel – Pathologic lung mucus



# Future plans

> To complete the analysis with the lung mucus Mu<sup>3</sup>Gel:

- I. Measure of metal ions content (by ICP-MS) in the collected medium by the basolateral side (*thanks to prof. Pacella, Università degli studi di Roma «La Sapienza»*)
- II. Rheological Analysis by <u>Frequency Sweep Test</u> (a particularly useful test as it enables the viscoelastic properties of a sample to be determined as a function of timescale). Several parameters can be obtained, such as the Storage (Elastic) Modulus (G'), the Viscous (Loss) Modulus (G''), and the Complex Viscosity and <u>stiffness</u> (η\*) (thanks to Daniela Pacheco, CEO of Bac3Gel)
- Co-culture with the EpiAirway model will be set up with naive and/or differentiated THP-1 to observe the inflammatory response in the lung tissue environment

# Future plans

**IARC point** to be completed by May 2023:

- Point 6: the onset of chronic inflammation through long-term EpiAirway co-culture with monocyte / macrophage M0 THP-1, endothelium and fibroblasts
- Point 8: Analysis of receptor-signal transduction modifications. Possible targets for analysis: mesothelin, TGF-beta, HMGB1 / RAGE